

Quantitative Analysis of Reticular Fibers in CCl₄-Induced Liver Cirrhosis in Mice.

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Abstract

Background

Cirrhosis is the final stage of chronic liver disease. It is characterized by diffuse fibrosis and disruption of hepatic lobular architecture into abnormal cirrhotic nodules.

Aim of the work

This study was conducted to investigate the extent of reticular fibers deposition in induced liver cirrhosis and their resolution after cessation of the insult.

Materials and methods

Three groups of animals were used; control, treated and recovery groups. The treated group injected subcutaneously with CCl₄ for 16 weeks for induction of cirrhosis, and the recovery group were kept for two weeks without injection after the 16 weeks of CCl₄ treatment.

Results

Our results indicated that reticular fibers increased in liver cirrhosis and decreased with fibers resolution in mice.

Conclusion

Reticular fibers are involved in the process of fibrogenesis in mice and decreased in the resolution of fibers in the recovery of the model which can be considered in new studies for the treatment of liver cirrhosis.

Keywords: liver, cirrhosis, fibrosis, CCl₄.

Abbreviations; CCl₄: carbon tetrachloride, ECM: extracellular matrix, HSCs: hepatic stellate cells.

Introduction

Liver cirrhosis is the final outcome of chronic liver diseases. Hepatic fibrosis is a wound-healing response which leads to accumulation of fibers in the extracellular matrix (ECM). Activation of hepatic stellate cells (HSCs); main source of ECM is key event in the development of hepatic fibrosis (Wang *et al.*, 2011).

Studying the pathogenesis of liver cirrhosis can help more effective treatment options to develop. Animal models are used to study fibrogenesis

and different strategies for the treatment of liver cirrhosis (Jang *et al.*, 2008).

Evidence that fibrosis and even cirrhosis are reversible has intensified interest in understanding fibrosis resolution, and the type of fibers. This can be applied in new therapies to reverse liver cirrhosis (Iredale *et al.*, 1998, and Domitrović *et al.*, 2009).

Materials and methods

Animals:

60 adult male Balb/c mice; about 2 months old with average weight 35 gm,

were purchased from Assuit Experimental Animal Facility, Assuit University. Animals were housed in Sohag University Animal House with free access to water and chow. They were acclimatized to this environment for 5 days prior to the experiment. All procedures used in this experiment were approved with the local Ethics Committee of Sohag University, Faculty of Medicine. Animals were randomly divided into four groups:

Group I (control): 5 animals were injected subcutaneously with the vehicle; sunflower oil twice weekly for 16 weeks, and another 5 animals were kept without treatment for 16 weeks

Group II: 25 animals were subcutaneously injected with 20% CCl₄ (Sigma Aldrich Company, Germany) in a dose of 1 ml /Kg twice weekly for 16 weeks for induction of cirrhosis (Vanheule *et al.*, 2008).

Group III: 25 animals were treated with CCl₄ for 16 weeks in the same dose of the previous group for induction of cirrhosis, then kept without for additional two weeks.

Methods:

Histological studies:

Liver samples were taken for processing of paraffin sections. Formalin fixed paraffin embedded section were used for detection of reticular fibers by silver stain according to *Bancroft et al.* (2013).

Morphometric studies

The light microscope Leica ICC50 Wetzlar (Germany) at the Histology Department, Faculty of Medicine, Sohag University was used, ten high power fields (x400) for each case in all groups were examined and analysis of each field was done using Image J software (version 1.46r) to detect the percentage areas of reticular fibers.

Statistical analysis

Paired sample Student *t*-test was used to analyze the data by using SPSS program (version 16.00; SPSS Inc., Chicago, Illinois, USA) with a statistical significance of $P < 0.05$. Data were expressed as mean \pm standard error (SEM).

Results

The reticular fibers in the control group appeared around the central veins, in the portal areas and in between the hepatic plates surrounding the sinusoids. In group II, reticular fibers significantly increased as compared to that in group I with formation of fibrous septa completely surrounding the hepatic lobules and the cirrhotic nodules. In group III, reticular fibers significantly decreased compared to that in group II; with fibrous septa partially surrounding the hepatic lobules (Fig. 1).

The mean percentage areas of reticular in different groups of the experiment were summarized in table (1), and Fig. (2).

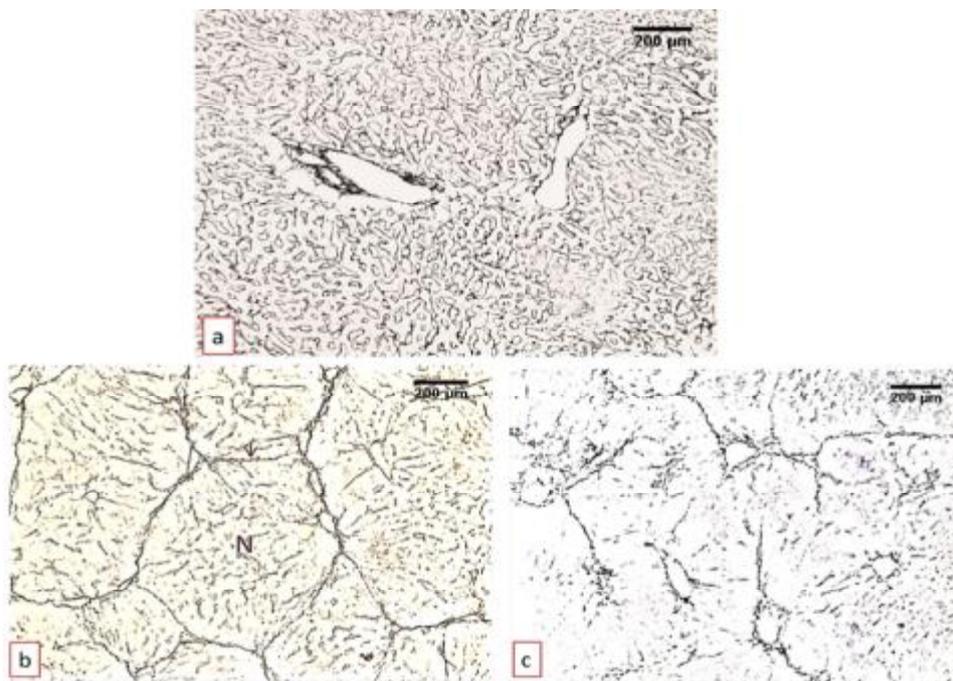


Fig. (1): Photomicrographs of liver sections from animals of; (a) group I showing reticular framework surrounding the central vein, in the portal area and around hepatic plates of hepatocytes.(b)group II showing numerous reticular fibers with complete septa surrounds the hepatic lobules and the cirrhotic nodules(N) and collapse of the fine fibers inbetween the parenchymal cells.(c) groupIII showing few reticular fibers around the central veins and in the portal areas with incomplete septae and inbetween the parenchymal cells (Silver stain).

Table (1): The mean percentage area of reticular fibers

Group	Reticular fibers percentage area. mean(\pm SEM)
Group I	10.92% (\pm 0.79)
Group II	14.26% (\pm 0.64) *
Group III	6.63% (\pm 0.60)*#

* Significant as compared to group IA (P value<0.05), # significant as compared to group II (P value<0.05)

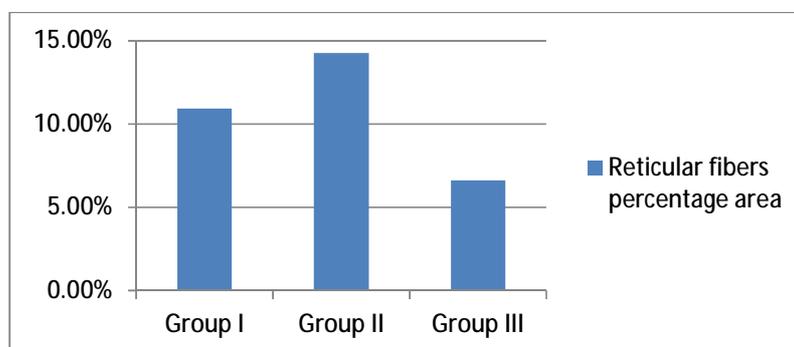


Fig.(2): The mean percentage area of reticular fibers.

Discussion

Hepatic fibrosis is caused by increased synthesis of ECM, produced by HSCs. (Wang *et al.*, 2011). In our study, reticular fibrous tissue significantly increased with disturbed architecture and formation of cirrhotic nodules. Similar findings were observed in previous studies Zaki *et al.* (2011) and Li *et al.* (2016) due to increased synthesis of reticular fibers by activated HSCs.

In our studies, we found that, 2 weeks after cessation of CCl₄ administration; there was significant reduction of reticular fibers; being demonstrated only in incomplete septae and in between the parenchymal cells. The reticular fibers in this group were less than the control due to the change of the type and characters of the extracellular matrix with collagen type I replacing the reticular fibers Zaki *et al.* (2011).

Our results were confirmed by previous reports. They proved that the key events in the resolution of liver fibrosis include decreased HSCs activation and increased degradation of fibers (Woessner, 1991). Other reports reported that hepatic myofibroblasts derived from HSCs undergo apoptosis during the spontaneous regression of liver fibrosis induced by CCl₄ (Iredale *et al.*, 1998). In contrast to our results, Domitrović *et al.*, (2009) showed persistence of fibrosis in their model 2 weeks after withdrawal of CCl₄. Kang *et al.*, (2005) also reported persistent fibrosis in their model after withdrawal of the insult with change of the localization of myofibroblast; being demonstrated only around the cirrhotic nodules.

In conclusion; reticular fibers deposition increase in a CCl₄ induced

model of liver cirrhosis in mice which can be applied in new strategies for the treatment of liver cirrhosis.

References

1. Domitrović R, Jakovac H, Tomac J, Sain I. (2009): Liver fibrosis in mice induced by carbon tetrachloride and its reversion by luteolin. *Toxicology and Applied Pharmacology*; 241: 311–321.
2. Iredale, J.P., Benyon, R.C., Pickering, J., McCullen, M., Northrop, M., Pawley, S., Hovell, C., Arthur, M.J., (1998): Mechanisms of spontaneous resolution of rat liver fibrosis. Hepatic stellate cell apoptosis and reduced hepatic expression of metalloproteinase inhibitors. *J. Clin. Invest.* 102, 538–549.
3. Jang JH, Kang KJ, Kim YH, Kang YN, Lee IS. (2008): Reevaluation of Experimental Model of Hepatic Fibrosis Induced by Hepatotoxic Drugs: An Easy, Applicable, and Reproducible Model. *Transplantation Proceedings*, 40, 2700–2703.
4. Kang JS, Morimura K, Salim EI, Wanibuchi H, Yamaguchi S, Fukushima S. (2005): Persistence of liver cirrhosis in association with proliferation of nonparenchymal cells and altered location of alpha-smooth muscle actin-positive cells. *Toxicol Pathol.*; 33(3):329–35.
5. Vanheule E, Geerts AM, Huysse JV, Schelfhout D, Praet M, Vlierberghe HV, Vos M.D, Colle I (2008) : An intravital microscopic study of the hepatic microcirculation in cirrhotic mice models: relationship between fibrosis and angiogenesis. *Int. J. Exp. Path.* ;89, 419–432.
6. Wang H, Lafdil F, Wang, L, Yin S, Feng D and Gao B. (2011): Tissue inhibitor of metalloproteinase 1 (TIMP-1) deficiency exacerbates carbon tetrachloride-induced liver injury and fibrosis in mice:

- involvement of hepatocyte STAT3 in TIMP-1 production. Cell Biosci 1, 14.
7. Woessner JF (1991). Matrix metalloproteinases and their inhibitors in connective tissue remodelling. FASEB J. 5, 2145–2154.
 8. Zaki MM, Ataa HM, Shenouda HD, Yousef MM, and Ahmed NE(2011): Effect of mesenchymal stem cells administered by two different routes on experimentally induced liver fibrosis in rats Egypt J Histol 34: 780-789.
 9. Li Z, Wei W, Chen B1, Cai G1, Li X1, Wang P1, Tang J1, Dong W(2016):The Effect of rhCygb on CCl4-Induced Hepatic Fibrogenesis in Rat. Sci Rep. 23;6:23508.