

CD34 Expression as a marker of progression of chronic hepatitis C

Maha Mohamed Agamy¹, Ghada Mostafa Kamal¹, Eman Muhammad Salah ElDeen², Nagwa Sayed Ahmad³, Asmaa Naser¹ Mohammad, Sheren Farrag Mahmoud².

¹Department of Tropical Medicine and Gastroenterology, Sohag Faculty of Medicine, Sohag University, ² Department of pathology, Sohag Faculty of Medicine, Sohag University, ³ Department of Medical Biochemistry, Sohag Faculty of Medicine, Sohag University.

Abstract

Background

In Egypt, the rate of death/transplantation, decompensation and HCC in patients with compensated HCV cirrhosis is 4.58%, 6.37% and 3.36%, respectively. It is estimated that Egyptian patients with HCV cirrhosis had an annual incidence of HCC with 5.3%. CD34 is a 110-kDa transmembrane glycoprotein present on leukemic cells, endothelial cells and stem cells). CD34 is preferentially expressed on the surface of regenerating or migrating endothelial cells and is a marker of proliferating endothelial cells in the growing sprouts during angiogenesis.

Aim of the work: The aim of the present study was to analyse the correlation between CD34 and the clinical severity of CHC.

Methods: A total of 70 adult patients with chronic hepatitis C infection in various stages with no evidence of cirrhosis, were recruited for the study. We studied the expression of CD34 in liver specimens from chronic HCV infected patients using a computer-based analysis of immunohistochemical staining and confirmed it by Western Blot.

Results: CD34 significantly related with AST and ALT levels. Also, it is significantly related with alpha-fetoprotein and prothrombin concentration. Also, There was a significant relation between CD34 and fibrosis stage.

Conclusion: A direct correlation existed between endothelial cell markers CD34 expression and the progression of fibrosis in chronic hepatitis C. The immunohistochemical methods of this molecule can be important clue in future prognostic strategies.

Introduction

Worldwide, more than 170 million people are infected with hepatitis C. In Egypt the prevalence of hepatitis C infection is considered the highest in the world [1]. In Egypt, the rate of death/transplantation, decompensation and HCC in patients with compensated HCV cirrhosis are 4.58%, 6.37% and 3.36%, respectively [2]. It is estimated that Egyptian patients with HCV cirrhosis had an annual incidence of HCC with 5.3% [3].

CD 34 is a cell surface, sialomucin-like glycoprotein expressed on hematopoietic progenitor cells, normal vascular endothelium and fibroblasts [4]. CD34 is an endothelial antigen used to highlight the density of vessels as a direct marker of the degree of neo-angiogenesis in tumors [5] and to distinguish well-differentiated HCC from non-neoplastic liver [6]. Understanding the process of angiogenesis might suggest an effective therapeutic target that would prevent disease progression. Data pertaining to

the assessment of angiogenesis in the liver tissue of CHC patients are scarce [6].

Angiogenesis is an integral part of tumor progression, it has also been observed in different inflammatory, fibrotic, and ischemic diseases [9]. Chronic liver diseases (CLDs) do not represent an exception to this rule. The suggestion that angiogenesis may significantly contribute to fibrogenesis and disease progression relies first on the fact that vascular remodeling is a common finding in human liver with advanced fibrosis, irrespective of etiology [6,8]. Capillary structures formed by ECs in inflamed portal tracts have been observed both in chronic hepatitis C (CHC) and chronic hepatitis B (CHB) [9,10]. Evidence of angiogenesis was significantly more frequent in HCV positive patients compared with HBV-positive patients or controls [9-11]. The aim of the present study was to analyse the correlation between CD34 and the clinical severity of CHC

Material and methods

70 patients with chronic hepatitis C had included in our study, all patients had undergone liver biopsy under ultrasonographic guide at the Department of Tropical Medicine And Gastroenterology of Sohag university hospitals, Sohag, Egypt.

The diagnosis of chronic HCV will be based on positive HCV antibody by ELISA test and HCV RNA by PCR for more than 6 months. The study was approved by the Ethical Committee of the Medical University of Sohag; Informed consent was obtained from all patients.

Histopathological assessment:

Seventy liver biopsies were included in the study and submitted

to histopathological examination to assess both the grade and the stage of chronic viral hepatitis, using the METAVIR classification system [12], which evaluates fibrosis stage in a scale from F0 to F4. Necro-inflammatory activity will be scored with a scale from A0 to A3.

Immunohistochemistry

CD34: For micro vessel density (MVD) assessment, we first identified „hot spots“ (areas with the highest micro vessel concentration) by scanning the section at low magnification (40x) using Olympus microscope CX21FS1, then we counted the number of positive vessels in three hot spots at a magnification of 200x, single immunoreactive endothelial cell, or endothelial cell clusters separate from other microvessels, were counted as a vessel (Weidner, 1995). The presence of blood cells or fibrin without any detectable endothelial cells was not sufficient to define a microvessel. Vessels with muscular walls were not counted (Svagzdys et al., 2009). The mean microvessel density for CD34 was calculated as the mean value of the vessel count in the three fields (Trojan et al., 2004).

Statistical analysis:

Data was analyzed using SPSS computer program version 22.0. Quantitative data was expressed as means \pm standard deviation, median and range. One-Way ANOVA test was used for normally distributed data. Spearman's correlation was used for testing of correlation between different quantitative variables. Chi-Square test was used for comparison between qualitative variables. P-values 0.05 were considered statistically significant.

Results

From September 2014 to august 2016, 70 adult patients with chronic hepatitis C infection had fulfilled our inclusion criteria and were included in this study. Their ages ranged from 20-59 years (40.5 ± 11.4 SD). 42 males, 28 females. Males and female were comparable according to age [p value 0.457]. Fibrosis stage of the patients using METAVIR classification system was done. The result were represented in table [1]

Fibrosis stage	Number	percentage
F1	28	28 (40%)
F2	22	22 (31.4%)
F3	20	20 (28.6%)

CD34 significantly related with AST and ALT levels. Also, it is significantly related with alpha-fetoprotein and prothrombin concentration.

Table [2] Correlation between CD34 and laboratory parameters of the studied patients (n= 70).

Group	R	P-value
HB (g/dl)	-0.056	0.647
RBCS (x1,000,000/mm3)	0.032	0.792
WBCS (x1000/mm3)	-0.181	0.133
Platelets (x1,000/mm3)	-0.164	0.175
ALT (IU/l)	0.264	0.027*
AST (IU/l)	0.295	0.013*
Bilirubin (mg/dl)	0.034	0.783
Albumin (mg/dl)	-0.189	0.118
Prothrombin time (sec)	0.214	0.075
Prothrombin concentration	-0.279	0.02*
Alpha fetoprotein	0.332	0.005*
HCV PCR	-0.022	0.860

There was asignificant relation between CD34 and fibrosis stage [p value=0.05].

Table [3]: Relation between fibrosis stage and laboratory data of the studied patients

Parameter	Fibrosis stage			P-value
	F1 (N= 28)	F2 (N= 22)	F3 (N= 20)	
HB (g/dl) Mean± S.D.	14.4 ± 1.5	14.1 ± 2.1	13.9 ± 1.8	0.640**
RBCS (x1,000,000/mm ³) Mean± S.D.	5.5 ± 0.9	5.1 ± 0.7	5.2 ± 0.8	0.305*
WBCS (x1000/mm ³) Mean± S.D.	6.9 ± 2.2	7.4 ± 3.5	5.7 ± 1.5	0.142*
Platelets (x1,000/mm ³) Mean± S.D.	244.3 ± 62.1	257.9 ± 65.6	201.6 ± 55.9	0.011**
ALT (IU/l) Mean± S.D.	46.2 ± 81.3	41.8 ± 26.4	73.2 ± 45.9	0.001*
AST (IU/l) Mean± S.D.	33.6 ± 28.8	41.7 ± 19.9	61.9 ± 32.9	< 0.001*
Bilirubin (mg) Mean± S.D.	0.7 ± 0.3	1.6 ± 3.7	0.7 ± 0.3	0.582*
Serum albumin (mg/dl) Mean± S.D.	4.2 ± 0.7	4.1 ± 0.4	4.01 ± 0.3	0.605*
Prothrombin time (sec) Mean± S.D.	11.9 ± 2.04	11.6 ± 0.9	12.6 ± 0.8	0.001*
Prothrombin concentration Mean± S.D.	93.3 ± 12.6	96.6 ± 9.3	86.4 ± 7.6	< 0.001*
Alpha fetoprotein Mean± S.D.	3.05 ± 1.3	5.01 ± 5.1	25.6 ± 38.4	0.001*
HCV PCR Low viremia (%) Moderate viremia(%) High viremia (%)	0 (0.0%) 8 (28.6%) 20(71.4%)	1 (4.5%) 4 (18.2%) 17 (77.3%)	3 (15%) 3 (15%) 14 (70%)	0.205***
CD34 levelMean± S.D.	15.7 ± 6.6	16.6 ± 7.3	26.1 ± 16.3	0.05*

Discussion

In the present study, there was a significant relation between CD34 and fibrosis stage. These observations indicate that angiogenesis is particularly linked to HCV infection, suggesting a possible contribution to HCV-related liver oncogenesis. Many studies reported that CD34 expression, which is a marker of neovascularization, was found in the periportal area of lobules and increased in parallel with the fibrosis stage. [13, 14].

Other studies reported that in CHC, new CD34 vessels are found in areas of lymphoid neogenesis which occurs in response to local cytokines in the portal tract to form expanded portal-associated lymphoid tissue [9, 16, and 17].

This observation additionally supports an interaction between the inflammatory activity and neoangiogenesis. It is possible that activated cytotoxic T cells may up-regulate the expression of vascular adhesion molecules, which enhance angiogenesis [15, 16]. The new vessels are an integral part of tissue remodeling that accompanies chronic inflammation and provide a portal of entry for the continuing recruitment of inflammatory cells. They also deliver nutrients and oxygen to areas of chronic inflammation that are relatively hypoxic [16]. CHC patients present a proangiogenic profile of angiogenesis soluble markers [17, 18]. CD34 is increased in viral hepatitis and their concentrations could be valuable markers of the evolution of liver inflammation, disease progression, and response to therapy [18, 19].

Conclusion

A direct correlation existed between endothelial cell markers CD34

expression and the progression of fibrosis in chronic hepatitis C. The immunohistochemical methods of this molecule can be an important clue in future prognostic strategies.

References

- 1- Blach S, Zeuzem S, Manns M, et al. Global prevalence and genotype distribution of hepatitis C virus infection in 2015: a modelling study. *Lancet Gastroenterol Hepatol.* 2016;2(3):161–176.
- 2- Alazawi W, Cunningham M, Dearden J, Foster GR. Systematic review: outcome of compensated cirrhosis due to chronic hepatitis C infection. *Aliment Pharmacol Ther.* 2010;32(3):344–355.
- 3- Eltabbakh M, Zaghla H, Abdel-Razek W, et al. Utility and cost effectiveness of screening for hepatocellular carcinoma in a resource limited setting. *Med Oncol.* 2015;32(4):432–437.
- 4- L.C. Strauss, S.D. Rowley, V.F. LaRussa, S.J. Sharkis, R.K. Stuart, C.I. Civin, et al., Antigenic analysis of hematopoiesis. Characterization of My-10 antigen expression by normal lymphohematopoietic progenitor cells, *Exp. Hematol.* 14 (9) (1986) 878–886.
- 5- F. Tanaka, Y. Otake, K. Yanagihara, Y. Kawano, R. Miyahara, M. Li, et al., Evaluation of angiogenesis in non-small cell lung cancer. Comparison between anti-CD34 and anti-CD105 antibody, *Clin. Cancer Res.* 7 (2001) 3410–3415.
- 6- M. Fernandez, D. Semela, J. Bruix J, I. Colle, M. Pinzani, J. Bosch, Angiogenesis in liver disease, *J. Hepatol.* 50 (2009) 604–620.
- 7- Chaparro M, Sanz-Cameno P, Trapero-Marugan M, Garcia-Buey L, Moreno-Otero R. Mechanisms of angiogenesis in chronic inflammatory liver disease. *Ann Hepatol* 2007; 6:208–213.

- 8- Gabriel A, Kukla M, Wilk M, et al. Angiogenesis in chronic hepatitis C is associated with inflammatory activity grade and fibrosis stage. *Pathol Res Pract* 2009; 205:758–764.
- 9- Mazzanti R, Messerini L, Monsacchi L, et al. CVH induced by hepatitis C but not hepatitis B virus infection correlates with increased liver angiogenesis. *Hepatology* 1997;25:229–234.
10. Messerini L, Novelli L, Comin CE. Microvessel density and clinicopathological characteristics in hepatitis C virus and hepatitis B virus related hepatocellular carcinoma. *J Clin Pathol* 2004;57:867–871.
11. Ohmori S, Shiraki K, Sugimoto K, et al. High expression of CD34-positive sinusoidal endothelial cells is risk factor for hepatocellular carcinoma in patients with HCV-associated chronic liver diseases. *Hum Pathol* 2001; 32:1363–1370.
- 12- Pinzani M, Rombouts K, Colagrande S. Fibrosis in chronic liver diseases: diagnosis and management. *J Hepatol* 2005; 42(Suppl): S22–36.
- 13- Gabriel A, Kukla M, Wilk M, et al. Angiogenesis in chronic hepatitis C is associated with inflammatory activity grade and fibrosis stage. *Pathol Res Pract* 2009; 205:758–764.
14. Kukla M, Gabriel A, Sabat D, et al. Association between liver steatosis and angiogenesis in chronic hepatitis C. *Pol J Pathol* 2010;3:154–160.
- 15- Jain RK. Molecular regulation of vessel maturation. *Nat Med* 2003; 9:685–693.
- 16- Lai WK, Adams DH. Angiogenesis and chronic inflammation; the potential for novel therapeutic approaches in chronic liver disease. *J Hepatol* 2005;45:7–11.
- 17- Girard JP, Springer TA. High endothelial venules (HEVs): specialized endothelium for lymphocyte migration. *Immunol Today* 1995;16:449–457.
- 18- Salcedo X, Medina J, Sanz-Cameno P, et al. The potential of angiogenesis soluble markers in chronic hepatitis C. *Hepatology* 2005;42:696–701.
- 19- Salcedo X, Medina J, Sanz-Cameno P. Review article: angiogenesis soluble factors as liver disease markers. *Aliment Pharmacol Ther* 2005;22:23–30.